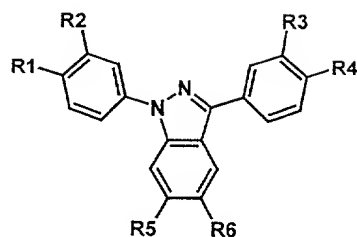


Claims:

1. Compounds represented by formula (I)



(Formula (I))

wherein

one of the radicals R^1 or R^2 and one of the radicals R^3 or R^4 is hydrogen and the other is independently $-\text{COOH}$, $-\text{COOR}^7$, $-\text{CONH}_2$, $-\text{CONH}(\text{CH}_2)_n\text{OH}$, wherein $n = 2 - 8$, $-\text{CONR}^8\text{R}^9$, $-\text{CH}_2\text{OH}$, $-\text{CH}_2\text{NH}_2$, $-\text{NO}_2$, $\text{NR}^{10}\text{R}^{11}$, NHCOR^{12} , Cl , Br , F , $-\text{CF}_3$, $\text{O}(\text{C}_1 - \text{C}_4)\text{-alkyl}$, which could be substituted by methyl or phenyl at any of the carbons $\text{C}_1 - \text{C}_4$, $-\text{N}=\text{C}=\text{O}$, $\text{N}=\text{C}=\text{S}$, $-\text{SO}_3\text{H}$, $-\text{SO}_2\text{NH}(\text{CH}_2)_n\text{NH}_2$, $(\text{C}_1 - \text{C}_4)$ alkyl, $(\text{C}_1 - \text{C}_{16})\text{-alkyl}$ substituted at the terminal carbon with $-\text{COOH}$, $-\text{COOR}^7$, $-\text{CONH}_2$, $-\text{CONR}^8\text{R}^9$, $-\text{CONH}(\text{CH}_2)_n\text{OH}$, wherein $n = 2 - 8$, $-\text{CH}_2\text{OH}$, $-\text{CH}_2\text{NH}_2$, $-\text{N}=\text{C}=\text{O}$, $\text{N}=\text{C}=\text{S}$, $-\text{SO}_3\text{H}$, $-\text{SO}_2\text{NH}(\text{CH}_2)_n\text{NH}_2$, $-\text{CONH}(\text{CH}_2)_n\text{NH}_2$, wherein $n = 2 - 8$, and the NH_2 -group could also be substituted by $(\text{C}_1 - \text{C}_4)$ alkyl or a commonly used amino protecting group such as *tert*-butyloxycarbonyl, 9-fluorenylmethoxycarbonyl, phthalimido, trifluoroacetamido, methoxycarbonyl, ethoxycarbonyl, benzyloxycarbonyl, allyloxycarbonyl, 2,2,2-trichloroethoxycarbonyl, 2-(trimethylsilyl)ethoxycarbonyl,

and one of the radicals R^5 or R^6 is hydrogen and the other is hydrogen, halogen, $\text{O}(\text{C}_1 - \text{C}_4)\text{-alkyl}$ which could be substituted by methyl or phenyl at any of the carbons $\text{C}_1 - \text{C}_4$, $-\text{NO}_2$, $\text{NR}^{10}\text{R}^{11}$, NHCOR^{12} , $(\text{C}_1 - \text{C}_4)$ alkyl,

(C₁-C₁₆)-alkyl substituted at the terminal carbon with -COOH, -COOR⁷, -CONH₂, -CONR⁸R⁹, -CONH(CH₂)_nOH, wherein n = 2 – 8, -CH₂OH, -CH₂NH₂, -N=C=O, N=C=S, -SO₃H, -SO₂NH(CH₂)_nNH₂, -CONH(CH₂)_nNH₂, wherein n = 2 – 8, and the NH₂-group could also be substituted by (C₁-C₄) alkyl or a commonly used amino protecting group,

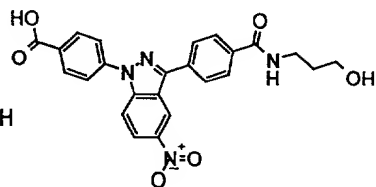
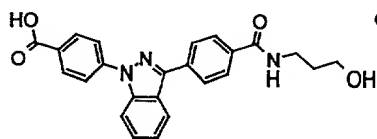
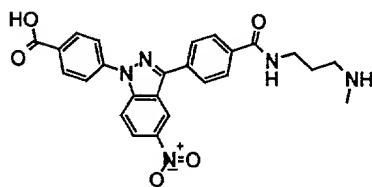
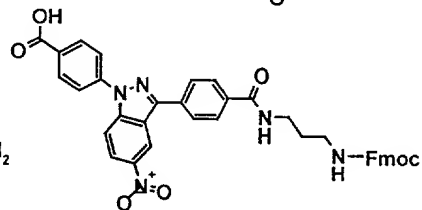
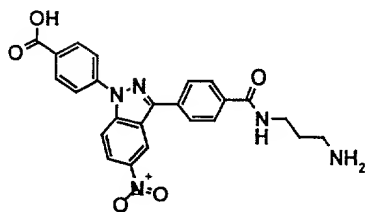
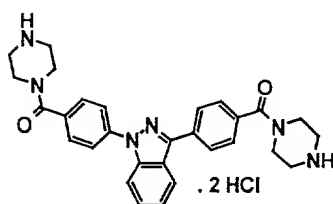
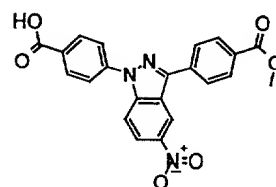
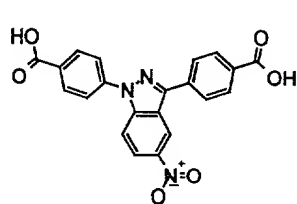
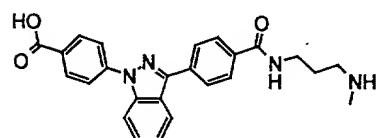
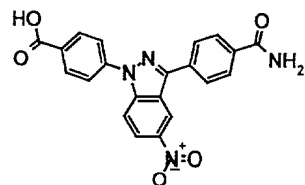
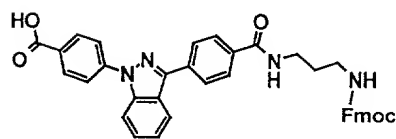
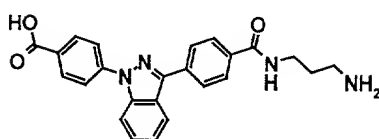
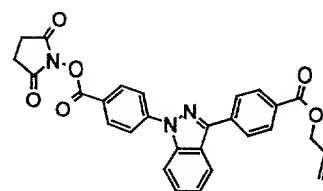
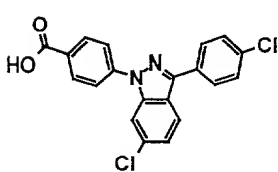
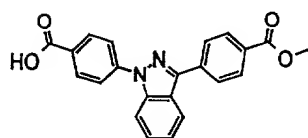
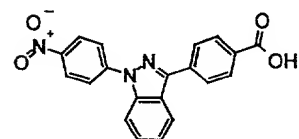
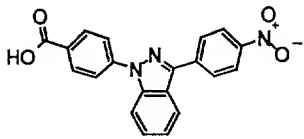
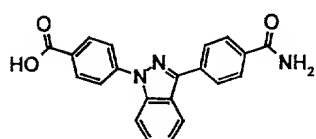
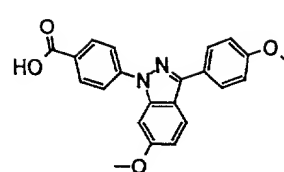
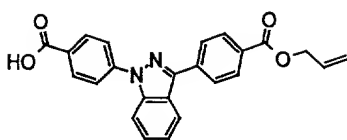
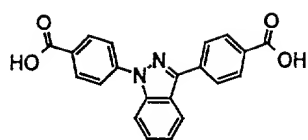
R⁷ is a commonly used carboxyl protecting or carboxyl activating group

R⁸ or R⁹ is hydrogen and the other is lower alkyl (C₁-C₄), phenyl, benzyl, or R⁸ and R⁹ are part of a 5 or 6 membered ring.

R¹⁰ and R¹¹ are independently hydrogen or (C₁-C₄)alkyl

R¹² is (C₁-C₁₀)alkyl, phenyl, which both can be substituted by (C₁-C₄) alkyl, protected amino group or halogen.

2. Compounds according to claim 1 represented by the following structures:



3. Compounds represented by formula (II – III)

A-B-D-C-D'-E (Formula (II))

A-B-D-E-D'-C (Formula (III))

wherein

A is a solid support selected from standard materials applied in solid phase and solution phase organic chemistry.

B is a linker allowing cleavage of fluorescent conjugates of formula (II-III) for liberation of the D-C-D'-E or D-E-D'-C fragment, respectively.

C is a compound selected from formula (I)

D and D' are independently a bond or a spacer selected from α,ω -diamino-alkanes, diaminocyclohexyl, bis-(aminomethyl)-substituted phenyl, α -amino- ω -hydroxy-alkanes, alkylamines, cyclic alkylamines or cyclic alkyldiamines or amino acids without or with additional functionality in the side chain.

E is the molecule to be investigated.

4. Compounds according to claim 3 wherein

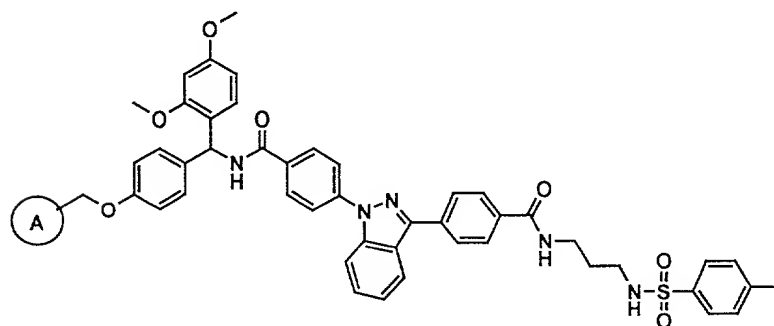
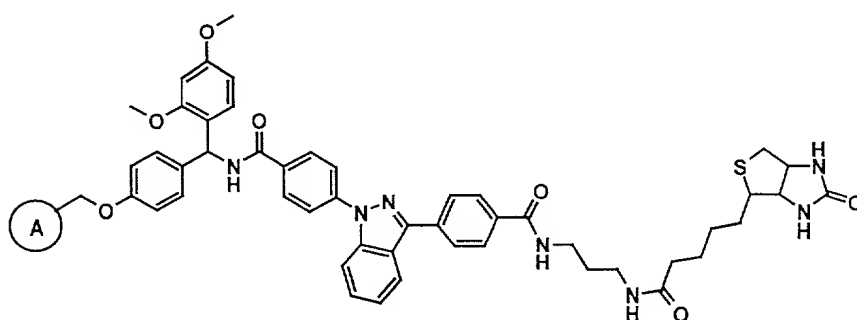
A is selected from functionalized polystyrene based resins, polyacrylamide based polymers, polystyrene / polydimethylacrylamide composites, PEGA resins, polystyrene-polyoxyethylene based supports, Tentagel, PEG-polystyrene graft polymeric supports, glass surfaces, functionalized surfaces, materials grafted with functionalized surfaces, or polyethyleneglycol.

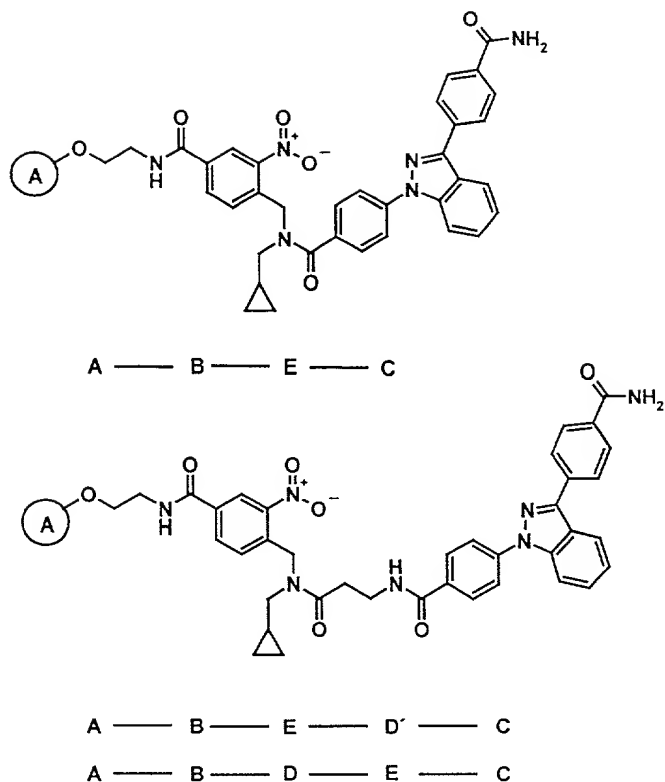
B is selected from benzyl, benzhydryl, benzhydryliden, trityl, xanthenyl, benzoin, silicon, or allyl based linkers.

C is a compound selected from formula (I)

E is a low molecular weight compound, a peptide, a protein, a carbohydrate, a nucleic acid, or a lipid containing a functional group for conjugate formation

5. Compounds according to claim 3 represented by the following structures:





6. Compounds represented by formula (IV):

E-D'-C (Formula (IV))

wherein

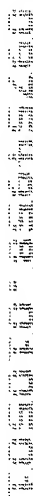
E is the molecule to be investigated

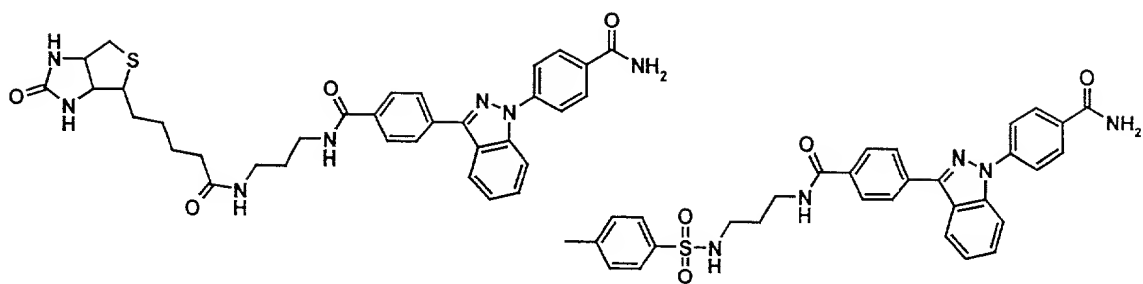
D' is a bond or a spacer selected from α,ω -diamino-alkanes, diaminocyclohexyl, bis-(aminomethyl)-substituted phenyl, α -amino- ω -hydroxy-alkanes, alkylamines, cyclic

alkylamines, cyclic alkyldiamines or amino acids without or with additional functionality in the side chain

C is a compound selected from formula (I)

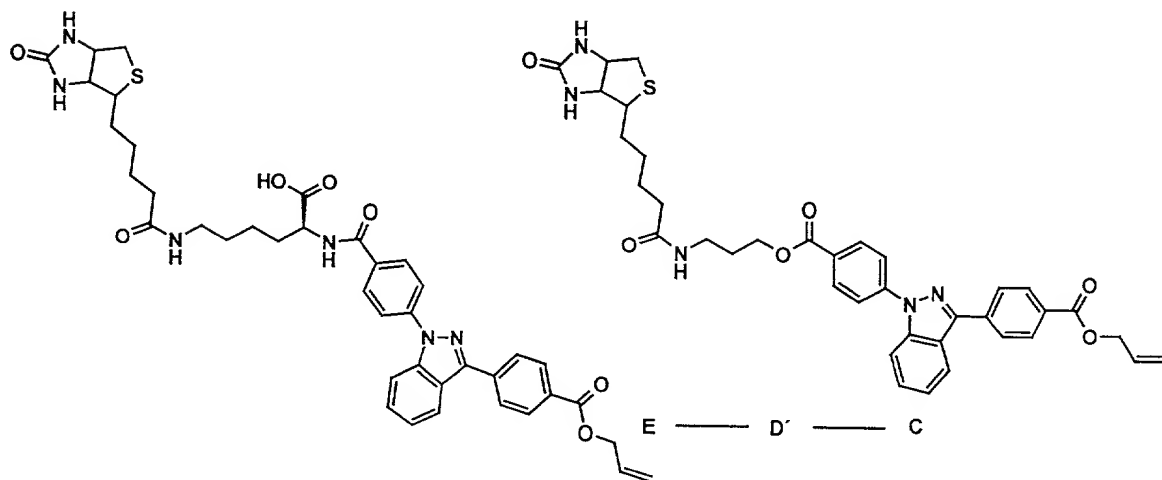
7. Compounds according to claim 6 represented by the following structures:





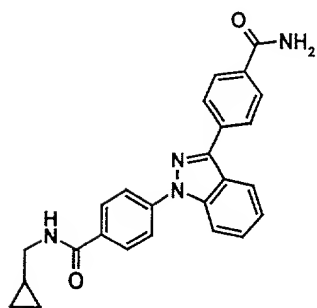
E — D' — C

E — D' — C

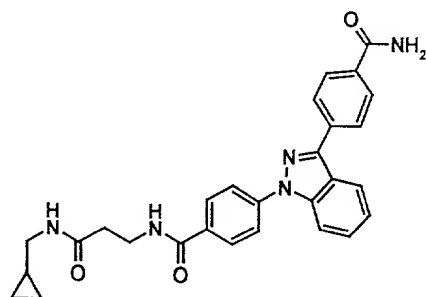


E — D' — C

E — D' — C



E — C



E — D' — C

8. A method for identification of an interaction between an AIDA labelled molecule and a binding molecule in homogeneous solution wherein the method comprises the following steps:

Step 1A: Providing an AIDA labelled molecule selected from formula (IV)

Step 1B: Admixing the AIDA-labelled molecule of formula (IV) with a binding molecule; and then

Step 1C: selectively detecting a binding event with the AIDA-labelled molecule described in Step 1B and the binding molecule by methods of fluorescence spectroscopy.

9. Method according to claim 8 wherein the methods of fluorescence spectroscopy are measurements of

- Increase of fluorescence anisotropy/polarisation of AIDA emission in continuous wave = prompt = steady state fluorometers,
- Increase of rotational correlation time in time-resolved fluorescence equipments
- Increase in translation diffusion time in single molecule fluorescence experiments determined from autocorrelation calculations on the time trace of fluorescence fluctuations,
- Increase or decrease of AIDA fluorescence emission in the wavelength range between 350 and 700 nm with excitation wavelengths in the range between 300 and 400 nm,
- Fluorescence resonance energy transfer (donor quenching or acceptor sensitisation) from excited tryptophan (donor) in the binding molecule which in this case is a peptide or protein to the AIDA dye (acceptor) in the molecule of the conjugate,
- Fluorescence resonance energy transfer (donor quenching or acceptor sensitisation) from the excited AIDA dye in the conjugate molecule (donor) to a fluorescent label (acceptor) of the binding molecule which in this case can comprise any compound class.

10. A method for identification of an interaction between an AIDA labelled molecule on the solid support which is conventionally used in solid phase organic chemistry and a binding molecule in homogeneous solution containing the solid support wherein the method comprises the following steps:

Step 2A: Providing an AIDA labelled molecule as conjugate of formula (II or III)

Step 2B: Admixing the AIDA-labelled molecule as conjugate of formula (II or III) with a binding molecule; and then

Step 2C: selectively detecting a binding event with the AIDA-labelled molecule described in Step 2B and the binding molecule by methods used in fluorescence spectroscopy resulting in a quantitative signal providing a means to identify the AIDA-linked molecule with the highest binding affinity to the binding molecule,

Step 2D: Isolation of the solid support containing the identified AIDA-molecule represented by formula (II or III)

Step 2E: : Selectively detecting a binding event with the AIDA-labelled molecule described in Step 2D and the binding molecule by various methods used in fluorescence spectroscopy described in the procedure 1A-C.

11. Method according to claim 10 wherein the fluorescence spectroscopic methods in step 2C are

- Direct detection of binding of fluorescently labelled macromolecules to AIDA containing solid supports applying confocal microscopic and spectroscopic techniques
- measurement of enhancement of the change in molecular brightness by chemically linking AIDA to a second environmentally sensitive molecule as

commonly used in conventional fluorescence spectroscopy performed during the synthesis of the compound on the solid support,

- measurement of fluorescence resonance energy transfer: From AIDA to a suitable long wavelength dye which will thereby be sensitised using AIDA UV-excitation detected by change in molecular brightness at the emission wavelength of the long wavelength dye,
- measurement of fluorescence resonance energy transfer: Reduction of specific brightness of AIDA on the molecule linked to the solid support at 351nm excitation and 400 nm emission wavelengths,
- Detection of the change in quantum yield by measuring reduction or increase in molecular brightness by time-resolved single molecule spectroscopy.